

**Polygenic transmission and complex neurodevelopmental network for
attention deficit hyperactivity disorder:
Genome-wide association study of both common and rare variants**

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Abstract

Attention deficit hyperactivity disorder ADHD is a complex polygenic disorder. This study aimed to discover common and rare DNA variants associated with ADHD in a large homogeneous Han Chinese ADHD case-control sample. The sample comprised 1,040 cases and 963 controls. All cases met DSM-IV ADHD diagnostic criteria. We used the Affymetrix 6.0 array to assay both single nucleotide polymorphisms (SNPs) and copy number variants (CNVs). GWAS were performed using PLINK. SNP-heritability and SNP-genetic correlations with ADHD in Caucasians were estimated with genome-wide complex trait analysis (GCTA). Pathway analyses were performed using the Interval enrichment Test (INRICH), the Disease Association Protein-Protein Link Evaluator (DAPPLE), and the Genomic Regions Enrichment of Annotations Tool (GREAT). We did not find genome-wide significance for single SNPs but did find an increased burden of large, rare CNVs in the ADHD sample ($P = 0.038$). SNP-heritability was estimated to be 0.42 (standard error, 0.13, $P = 0.0017$) and the SNP-genetic correlation with European Ancestry ADHD samples was 0.39 (SE 0.15, $P = 0.0072$). The INRICH, DAPPLE, and GREAT analyses implicated several gene ontology cellular components, including neuron projections and synaptic components, which are consistent with a neurodevelopmental pathophysiology for ADHD. This study suggested the genetic architecture of ADHD comprises both common and rare variants. Some common causal variants are likely to be shared between Han Chinese and Caucasians. Complex neurodevelopmental networks may underlie ADHD's etiology.

INTRODUCTION

Attention-deficit hyperactivity disorder (ADHD) is a common behavioral disorder of childhood, affecting 3–6% of school-age children around the world [Faraone et al., 2003]. It has been viewed as a polygenic, multifactorial disorder. Both common and rare DNA variants contribute to its complex etiology [Poelmans et al., 2011; Stergiakouli et al., 2012; Williams et al., 2012].

Genome-wide association studies (GWAS) are hypothesis-free, interrogate all genes and regulatory regions of the genome and have the potential to discover novel risk genes. The first GWAS of ADHD performed by Neale et al. [2008] analyzed 438,784 SNPs in 909 Caucasian ADHD trios. Although none of the SNP association tests achieved genome-wide significance, the top-25 SNPs (based on P -value) implicated some interesting candidate genes, including cytoskeleton organizer DCLK1, extracellular matrix component SPOCK3,

cell-cell adhesion protein CDH13, as well as two potassium channel regulators KCNIP1 and KCNIP4. Using the same sample set, Lasky-Su et al. [2008] performed a quantitative genome-wide association analysis of ADHD symptoms. A high percentage (30/32, 94%) genes hit by the 58 SNPs with P values less than 10^{-5} were brain-expressed, including five related to transcription factors.

Meanwhile, Lesch et al. [2008] used independent DNA pools from 343 ADHD-affected adults and 304 controls for association analyses of the ADHD diagnostic phenotype. Of the 30 top-hit genes, seven were involved in cell adhesion/migration/neurogenesis (e.g., CDH13, ASTN2, CSMD2, ITGAE, ITGA11, CDH23, SDK2), two regulated synaptic plasticity (e.g., CTNNA2, KALRN), three were transcription factors (MYT1L, TFEB, SUPT3H), and one coded for a potassium channel (KCNC1) [Lesch et al., 2008].

Neale et al. [2010a] performed case-control analyses in 896 cases with DSM-IV ADHD and 2,455 controls. A consensus dataset of 1,033,244 SNPs was imputed (using the HapMap Phase III European CEU and TSI samples as the reference). No genome-wide significant associations were found. The most significant results implicated PRKG1, FLNC, TCERG1L, PPM1H, NXP1, CDH13, HK1, and HKDC1. Combining data from four ADHD GWAS projects,

Neale et al. [2010b] performed a meta-analysis in a sample of 2,064 trios, 896 cases, and 2,455 controls. Even with this much larger sample size, no genome-wide significant associations were found. One reason for this is that the samples were underpowered to estimate effect sizes of common variants of small effect. This has been confirmed by analyses that estimate the variance contributed by common variants all together. Genome-wide complex trait analysis (GCTA) [Yang et al., 2010, 2011; Lee et al., 2011] applied to ADHD samples (4,163 cases and 12,040 controls) from the Psychiatric Genomics Consortium, estimated SNP chip heritability to be 0.28 (SE 0.02; Psychiatric GWAS Consortium ADHD Group. Paper submitted for publication).

Copy number variations (CNVs) have also been implicated in the etiology of ADHD. Elia et al. [2010] found that inherited rare CNVs in an ADHD sample were significantly enriched for genes known to be important for psychological and neurological functions, including learning, behavior, synaptic transmission, and central nervous system development. Williams et al. [Williams et al., 2010, 2012] found an increased burden of large, rare CNVs and reported excess of chromosome 16p13.11 and 15q13.3 duplications and an overlap between CNVs reported for ADHD and autism spectrum disorders. Elia et al. [2012] further showed that CNVs affecting the metabotropic glutamate

receptor genes GRM5, GRM7, GRM8, and GRM1 were enriched across several independent samples.

In summary, although ADHD is acknowledged to be a genetic disorder, GWAS has not revealed any common SNP variants with genome-wide significance. This study used both common and rare variants, using polygenic and pathway analyses, to evaluate the genetic etiology of ADHD in a large homogenous Han Chinese case-control sample.

MATERIALS AND METHODS

Participants

One thousand and forty ADHD cases (876 boys, 84.2%) aged between 6 and 16 years [average (9.7 ± 2.4) years] were recruited from the Child and Adolescent Psychiatric Outpatient Department of the Sixth Hospital, Peking University. All cases met DSM-IV ADHD diagnostic criteria. A clinical diagnosis was first made by a senior child and adolescent psychiatrist based on the parent and teacher completed ADHD Rating Scale-IV (ADHD-RS-IV), and then confirmed by semi-structured interview with the parents and child using the Chinese version of the Clinical Diagnostic Interview Scale [Barkley, 1998; Yang et al., 2004]. Those with major neurological disorders (e.g., epilepsy), schizophrenia, pervasive development disorder, and mental retardation (IQ < 70) were excluded. The sample consists of 680 (65.4%) ADHD combined type and 360 (34.6%) inattentive type. The comorbidities included oppositional defiant disorder (ODD) in 380 patients (36.5%), conduct disorder in 58 (5.6%), and tic disorder in 167 (16.1%).

Nine hundred sixty three controls were students from local elementary schools, healthy blood donors from the Blood Center of the First Hospital, Peking University, and healthy volunteers from our institute. Six hundred and

eight were males (63.1%). The average age was 15.4 ± 8.8 ys. Parents or adults themselves completed the ADHD Rating Scale-IV (ADHD RS-IV) to exclude ADHD. Major psychiatric disorders, family history of psychosis, severe physical diseases, and substance abuse were also excluded according to a medical history report form. All the cases and controls were of Han Chinese decent.

The study was approved by the Institutional Review Board of the Peking University Health Science Center. After complete description of the study to the subjects, written informed consent was obtained from parents of the ADHD probands.

Genotyping

Both cases and controls were genotyped using the Affymetrix6.0 array at CapitalBio Ltd. (Beijing) using the standard Affymetrix protocol. Samples of cases and controls were added in equal proportion to each chip to avoid batch effects. The Affymetrix 6.0 array included 906,600 SNP probes and 946,000 CNV probes. The SNP genotypes were called with BIRDSEED v2, while CNVs were called with Genotyping Console (GTC) 4.0 using default parameters. A total of 2003 cases and controls passed the first stage sample control with call rates $>98\%$, no first or second-degree relative relationships, and genders consistent with site reports.

Data Quality Control and Statistical Analysis

Data quality control and association analysis were performed using PLINK 1.07 [Purcell et al., 2007, <http://pngu.mgh.harvard.edu/purcell/plink/>]. For inclusion of SNPs we required: call rate $>95\%$, MAF $>1\%$ and HWE P-value $>10^{-6}$. After data cleaning, there were 656,051 SNPs for the association analyses. To examine population stratification, we performed multi-dimensional scaling (MDS). In the pair-wise MDS plot for 10 dimensions, the majority of subjects were tightly clustered, suggesting no

substantial population stratification (SF1). Wethen conducted logistic regression to adjust the association P-value, using the 10 principal components from the MDS procedure as covariates.

CNV calling only included segments larger than 100 kb, spanning at least 10 consecutive, informative SNPs. Quality control for samples excluded 136 individuals (71 cases, 65 controls) who carried more than 40 apparent CNVs. Analysis focused on rare CNVs with frequency $<1\%$. We used the human reference sequence of NCBI Build 36.1 - hg18 to filter known segmental duplications. Known common CNVs defined by the Genome Structural Variation Consortium (http://projects.tcag.ca/variation/ng42m_cnv.php) and known gaps of at least 200 kb in the SNP array were also filtered. Burden analysis counted the number of total CNVs, deletions and duplications in cases and controls, calculated the CNV rate, as well as per cent of cases and controls that carried rare CNVs. The significance of CNV differences between cases and controls was assessed by permutation test with 50,000 replicates.

Polygenic analyses

To investigate the contribution of common SNPs to variation in liability to ADHD, we estimated the SNP heritability using GCTA [Yang et al., 2011]. A non-zero heritability is estimated if cases are genetically more similar to other cases than they are to controls [Lee et al., 2011]. We removed individuals such that no pair had genetic similarity relationship >0.05 (as this may inflate estimates unfairly), so that 1,010 cases and 917 controls remained. We used Caucasian samples from the Psychiatric Genomics Consortium for ADHD (4,163 ADHD cases and 12,040 controls) and a bivariate model of analysis [Lee et al., 2012b] to estimate the SNP-genetic correlation between Han Chinese and Caucasians for liability to ADHD. Since

the SNP frequencies differ between ethnic groups the additive genetic similarities between individuals i and j were estimated as

$$A_{ij} = \frac{1}{L} \sum_{l=1}^L (x_{il} - 2p_{is,l}) \frac{x_{jl} - 2p_{js,l}}{\sqrt{2P_{is,l} Q_{is,l} 2P_{js,l} Q_{js,l}}} \quad i \neq j$$

for the L SNPs with minor allele frequency >0.01 and imputation $R^2 > 0.6$ ($L = 917,066$), where i and s represent a population that individual i belong to and p and $q = 1 - p$ are allele frequencies of the first and other allele and x_{il} is the number of first alleles for the l th SNP in individual i . The analysis model include sex, cohort and 20 ancestry principal components are covariates.

Pathway analysis

To determine if any neurobiological pathways were implicated by our association signals, we input our top hit intervals from the SNP and CNV association analyses into Interval enrichment Test (INRICH [Lee et al., 2012a]). Associated intervals for SNPs included those with P -values $<10^{-4}$ after correcting for the MDS components. The SNP tagging function in PLINK was used to generate LD independent genomic intervals (tag r^2 : 0.2, tag kb: 1,000). We included CNV intervals that were more prevalent in cases than in controls with at least a trend difference of statistical significance ($P < 0.15$). We used the Gene Ontology (GO) nodes as our target gene sets. After size filtering, 5,237 target gene sets (nodes) each comprising at least three genes were examined. Interval overlap was limited to 20 kb up/downstream of a gene. The number of overlapping genes was recorded as $Real_i$. Ten thousand replicates generated random interval sets each matching to the number of associated intervals. The empirical gene-set P -value equals the per cent of replicates with at least $Real_i$ number of random intervals overlapping with genes in a target gene set. Bootstrapping-based re-sampling was used for multiple testing to correct the empirical gene-set P -value over all gene sets.

To explore potential physical interactions among proteins encoded in associated intervals, we used a second method for pathway analysis, that is, Disease Association Protein-Protein Link Evaluator (DAPPLE) [Rossin et al., 2011]. In consideration of the contributions of both common and rare variants to the etiology of ADHD, and that both might separately capture nodes in the ADHD pathogenesis network, we used the same genomic intervals for both SNPs and CNV that we used for INRICH. DAPPLE uses experimentally validated, protein-protein interaction databases to identify direct and indirect networks from associated proteins and scores network and protein connectivity. We built 10,000 random networks and compared these with the ADHD associated networks to determine if the connectivity of the ADHD networks and each seed protein was greater than expected by chance.

The third pathway analyses we used the Genomic Regions Enrichment of Annotations Tool (GREAT, [McLean et al., 2010]) to assess for enrichment of cis-regulatory regions. GREAT examines not only proximal but also distal regulatory regions up to 1 Mb upstream or downstream of transcription start sites. In addition to typical calculation of gene-based P -values for enrichment, GREAT computes a binomial test over genomic regions, which uses the fraction of the genome associated with each ontology term as the probability of selecting the term. This method explicitly accounts for the variability in length of gene regulatory domains, eliminating the bias that leads to false positive enrichments for distal regulatory regions.

RESULTS

Single Variant Analyses

The quantile-quantile (QQ) plot (SF2) for SNPs' association was almost completely diagonal. The lambda statistic (λ) was 1.02. The distribution of observed P -values did not deviate from

the distribution expected under the null hypothesis of no association. The corrected Manhattan plot is shown in SF3. The lowest P values were about 10^{-5} to 10^{-6} . The SNPs associated with P values of 10^{-5} or lower are listed in Supplementary Table SI. All hit genes were expressed in brain. Most of them were known to be involved in neurodevelopment (including cell adhesion, neuron migration, neurite outgrowth, neuronal morphogenesis, and synaptic plasticity: ITGA1, NYAP2, ADAM28, CNTN2, LRFN2, NTM, GJA1, FLRT2, PRKG1, PICK1, CAMK2G; glutamate receptor and transporter: GRIK4, GRM7, SLC38A1; and related transcription factors: PAPOLA, MED27, TAF2, ZNF516).

We included 3,460 rare CNVs (1,817 in cases and 1,643 in controls) in the analyses, with all segments intersecting with one or more genes (hg18). Burden analyses showed a significantly higher rate of rare CNVs (1.875% vs. 1.830%, ratio: 1.02, $P = 0.038$) and proportion of individuals carrying rare CNVs (55.8% vs. 51.2%, ratio: 1.09, $P = 0.026$) for the ADHD group than for controls. Association analyses found six regions nominally associated with ADHD ($P < 0.05$, with 50,000 permutation tests), though none of them survived genome-wide correction (Supplementary Table SII).

Polygenic Analyses

The estimate of the SNP-heritability calculated in the bivariate analysis was 0.42 (SE 0.13) for the Han Chinese sample ($h^2_{\text{SNP}_{\text{HC}}}$). A maximum likelihood ratio test of $H_0: h^2_{\text{SNP}} = 0$ is $P = 0.0017$. In the bivariate analysis the SNP-heritability for the European ancestry sample (h^2_{SNPEA}) was 0.28 (SE 0.02, $P = 0$), in close agreement (as expected) with the univariate estimate PGC Cross Disorder Group, paper in submission. The estimate of the SNP-genetic correlation between Chinese and

European samples ($r_{\text{g-SNP}}$) was 0.39 (SE 0.15, $P = 0.0072$).

Pathway Analyses

Interval enrichment tests of the most significantly associated SNPs found 23 pathways enriched for associated signals (Table I). Although none of these achieved significance after correcting for multiple comparisons, many implicated neurobiological functions potentially relevant to ADHD, e.g. neuron projection morphogenesis (ITGA1, GJA1), neuron migration (PRKG1, GJA1), endocytic vesicle membrane (PICK1, CAMK2G), synaptic transmission (PICK1, CAMK2G, SLC38A1, GRM7). Pathways related to transcription were observed, that is, transcription initiation from RNA polymerase II promoter (MED27, TAF2). Interval enrichment tests of CNVs found 9 pathway nominal significant at $P < 0.05$. None achieved significance after correcting for multiple comparisons (Table I). Most were related to transmembrane transport, including water, sodium and potassium ion transport.

DAPPLE identified 16 direct connections among proteins in 152 associated regions (Table II). Compared to 10,000 random networks, the associated network (SF4) is significantly enriched for direct connectivity (16 vs. 9.7, $P = 0.030$). The connected proteins formed six groups. Their functions involved cell adhesion/synaptic formation/plasticity, especially for glutamatergic synaptic plasticity, as well as related transcription factors. For each seed protein, taking the best of the direct and indirect scores and correcting for the number of tests as well as for the number of genes in one locus, we identified seven genes significant for connectivity to be candidate genes for future research: NCL ($P = 2 \times 10^{-4}$), KCNH7 ($P = 8 \times 10^{-4}$), NXP1 (1×10^{-3}), LANCL1 (6×10^{-3}), CNTNAP2 (9×10^{-3}), SV2C (1.2×10^{-2}), and PICK1 (4×10^{-2}).

Using the same set of associated SNPs and CNVs for GREAT analyses, we found

TABLE I.

Pathways Enriched for Associated SNPs and CNVs by INRICH Test

Target	Target Size	Interval no.	Emp. P	Cor. P	Gene list
Pathways enriched for associated SNPs^a					
GO: 0009268Response_to_pH	12	2	0.00009999	0.14917	ARSB, GJA1
GO: 0043403Skeletal_muscle_tissue_regeneration	9	2	0.00019998	0.192162	PLAU, GJA1
GO: 0048812Neuron_projection_morphogenesis	18	2	0.00029997	0.231354	ITGA1, GJA1
GO: 0007160Cell-matrix_adhesion	72	3	0.00049995	0.295941	VCL, ITGA1, BCL2L11
GO: 0005916Fascia_adherens	9	2	0.00079992	0.378724	VCL, GJA1
GO: 0006936Muscle_contraction	93	3	0.00109989	0.447111	VCL, ITGA1, GJA1
GO: 0030666Endocytic_vesicle_membrane	24	2	0.00269973	0.661668	PICK1, CAMK2G
GO: 0006367Transcription_initiation_from_RNA_polymerase_II_promoter	67	2	0.00379962	0.727854	MED27, TAF2
GO: 0005741Mitochondrial_outer_membrane	85	2	0.00679932	0.847231	GJA1, BCL2L11
GO: 0007229Integrin-mediated_signaling_pathway	58	2	0.010399	0.894421	ITGA1, ADAMDEC1
GO: 0005178Integrin_binding	64	2	0.0114989	0.907419	ITGA1, ADAMDEC1
GO: 0007268Synaptic_transmission	266	4	0.0121988	0.915217	PICK1, CAMK2G, SLC38A1, GRM7
GO: 0005764Lysosome	154	2	0.0157984	0.946011	ARSB, GJA1
GO: 0030165PDZ_domain_binding	54	2	0.019898	0.965807	GRM7, GJA1
GO: 0015293Symporter_activity	112	2	0.019898	0.965807	SLC38A1, SLC16A8
GO: 0005624Membrane_fraction	467	4	0.0243976	0.982004	ITGA1, SLC16A8, PSD3, BCL2L11
GO: 0001764Neuron_migration	59	2	0.0255974	0.983003	PRKG1, GJA1
GO: 0045121Membrane_raft	110	2	0.0256974	0.983203	ITGA1, GJA1
GO: 0006814Sodium_ion_transport	118	2	0.0265973	0.984003	SLC38A1, SCN9A
GO: 0005654Nucleoplasm	732	4	0.0214979	0.968806	MED27, CAMK2G, DSCC1, PAPOLA
GO: 0017124SH3_domain_binding	101	2	0.0357964	0.992402	BAIAP2L2, GJA1
GO: 0043234Protein_complex	152	2	0.0437956	0.995601	PICK1, VCL
GO: 0001701In_utero_embryonic_development	142	2	0.0459954	0.996601	GJA1, BCL2L11
Pathways enriched for associated CNVs^b					
GO: 0006833Water_transport	36	2	0.00549945	0.508498	AQP9, ADCY8
GO: 0005244 Voltage-gated_ion_channel_activity	149	4	0.0115988	0.734653	KCNH7, KCNQ1, SCN10A, SCN11A
GO: 0055085Transmembrane_transport	619	6	0.0125975	0.752849	KCNH7, SCN10A, KCNQ1, AQP9, ADCY8, SCN11A
GO: 0042493Response_to_drug	241	3	0.0164984	0.834233	USP47, CPS1, SCN11A
GO: 0005248 Voltage-gated_sodium_channel_activity	15	2	0.0172983	0.84883	SCN10A, SCN11A
GO: 0001518 Voltage-gated_sodium_channel_complex	12	2	0.0172983	0.84883	SCN10A, SCN11A
GO: 0006813Potassium_ion_transport	153	3	0.0392961	0.956809	KCNH7, KCNQ1, KCNT2, SCN11A
GO: 0006814Sodium_ion_transport	118	2	0.0258974	0.909018	SCN10A, SCN11A
GO: 0006811 Ion_transport	565	5	0.030397	0.928814	KCNH7, KCNQ1, KCNT2, SCN10A,

a With corrected P-value $<10^{-4}$.

b Including CNVs more in cases than in controls with $P < 0.15$.

significant enrichment for 6 GO Cellular Component terms after correcting for multiple comparisons (Table III). The six terms were from two GO branches and their child nodes (Fig. 1): synapse (15 genes hit, FDR Q -val: 0.0055; three child nodes were also significant: synapse part, synaptic membrane, and presynaptic membrane) and neuron projection (16 genes hit, FDR Q -val:

0.013; one child node was also significant: axon).

DISCUSSION

This GWAS of ADHD, comprising 1,040 cases and 963 controls, is the first performed in a homogeneous Han Chinese population. Although we did not find any genome-wide significant SNP or CNV variants, we did find significant

evidence for a polygenic SNP component and an increased burden of rare CNVs.

The significant SNP-heritability implies that common variants are associated with ADHD, but that our sample is underpowered to detect them at the stringent significance level imposed by the genome-wide burden of multiple testing. The SNP-heritability in Han Chinese was 0.42 (SE 0.15). Although the point estimate is higher than for the larger European ancestry sample from the PGCADHD, 0.28 (SE 0.02), its high standard error shows that the estimates are not significantly different. The estimate of the SNP-genetic correlation (r_{g-SNP}) was 0.39 (SE 0.15, $P \approx 0.0072$), which indicates that common SNP risk variants are shared by the Han Chinese and European Ancestry samples. To our knowledge, this is the first such correlation reported for any disease or disorder. The significant correlation indicates that ancient common variants associated with ADHD are shared between the ethnic groups. However, the point estimate of the SNP correlation between Han Chinese and European Ancestry samples is lower than between sub-samples of the European Ancestry cohort. Specifically, when the PGC-ADHD data was split into two sub-samples, the $h^2_{SNP,EA}$ estimates were 0.21 (SE 0.07) for the first sample and 0.41 (SE 0.03) for the other sample with a genetic correlation of 0.71 (SE 0.17) implying, as expected, more sharing of associated variants and/or higher linkage disequilibrium between causal variants and SNPs within than between ethnic populations.

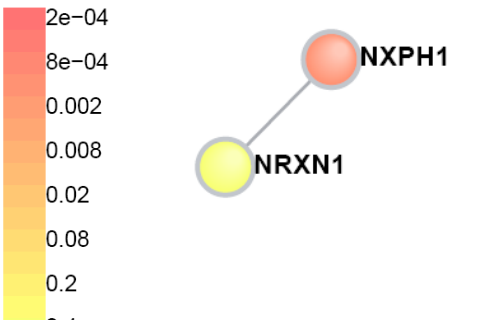
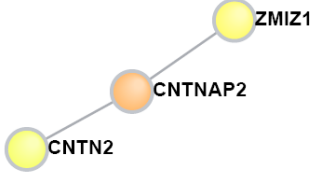
Despite the fact that no individual SNPs reached association at genome-wide significance, our most significant findings implicated genes participating in neurodevelopmental processes such as cell adhesion, neuron migration, neurite outgrowth, neuronal morphogenesis, and synaptic plasticity. Similar sets of genes were also suggested by

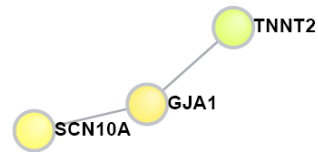
previous ADHD GWAS and a meta-analysis (see Supplementary Table SI). For example, PRKG1 was implicated by Neale et al. [2010a], ITGA1, CAMK2G, CAMK1D were implicated in the meta-analysis by Neale et al. [2010b], and ITGAE and ITGA11 were implicated by Lesch et al. [2008]. Some of our top genes code for glutamate receptors and transporters. The same genes and gene family members (GRM7, GRIK1) were reported in the quantitative GWAS by Lasky-Su et al. [2008], the meta-analysis by Neale et al. [2010b], and the genome-wide CNV study by Elia et al. [2012]. Some genes related to transcription (ZNF544, ZNF385D, ZNF423, ZNF516, ZNF75A, DMRT2, FHIT, FOXP1, and MEIS2) were also implicated by Lasky-Su et al. [2008] and by Neale et al. [2010b]'s meta-analysis.

Although not significant after correcting for multiple comparisons, the pathways revealed by the INRICH analyses of associated SNPs involved neurobiological functions consistent with the prior findings discussed above. For example, neuron projection morphogenesis and neuron migration pathways were implicated by genes encoding adhesion molecules (e.g., GJA1, ITGA1, and PRKG1). Neuron migration and axon guidance toward the target in the development of the nervous system involve interactions between molecules on the surface of the axon and those in the extra-cellular matrix [Tsiotra et al., 1993]. The endocytic vesicle membrane and synaptic transmission pathways involve glutamatergic synaptic function. The transcription related pathway is a ubiquitous biological process, if, as our findings suggest, it is implicated in ADHD's pathophysiology, any defects in the implicated transcription network must require other etiological factors to lead to a pathophysiologic state.

Because the GO "pathways" used by INRICH are based on bibliometric gene annotations rather than experimental

Table 2. Direct connections between proteins encoded by genes overlapped with potential associated SNPs and CNVs

Direct connection	Associated SNP/interval	Gene function*	Gene & pathway implicated in other studies of ADHD or psychiatric disorders
	<p><i>NXP1</i>: chr7: 8471930-8567358 <i>NRXN1</i>: chr2: 50226778-50323795</p>	<p><i>NXP1</i>: The encoded protein forms a very tight complex with alpha neurexins, a group of proteins that promote adhesion between dendrites and axons.</p> <p><i>NRXN1</i>: The encoded protein functions as cell adhesion molecules and receptors. May play a role in formation or maintenance of synaptic junctions.</p>	<p>Both <i>NXP1</i> and <i>NRXN1</i> were among the top hit genes (10^{-5}) in ADHD GWAS by Neale et al. (2010). CNVs of <i>NXP1</i> were found in ASD families (Salyakina and others 2011). CNVs of <i>NRXN1</i> were also observed in schizophrenia and autism (Doherty and others 2012).</p>
	<p><i>CNTN2</i>: rs2802837 <i>CNTNAP2</i>: chr7: 145653708-145657817 Chr7: 145755346-145813647 <i>ZMIZ1</i>: chr10: 80350172-80476638</p>	<p><i>CNTN2</i>: It is a neuronal membrane protein that functions as a cell adhesion molecule. It may play a role in the formation of axon connections in the developing nervous system.</p> <p><i>CNTNAP2</i>: This gene encodes a member of the neurexin family which functions as cell adhesion molecules and receptors. This protein is localized at the juxtaparanodes of myelinated axons, and mediates interactions between neurons and glia during nervous system development and is also involved in localization of potassium channels within differentiating axons.</p> <p><i>ZMIZ1</i>: The encoded protein regulates the activity of various transcription factors</p>	<p><i>CNTNAP2</i> has been implicated in multiple neurodevelopmental disorders, including Gilles de la Tourette syndrome, schizophrenia, epilepsy, autism, ADHD and mental retardation. <i>CNTNAP5</i> was among the top hit genes in ADHD GWAS by Neale, et al. (2010). <i>ZMIZ1</i> was among the top hit genes in Neale, et al., (2008). <u>Cell adhesion</u> molecule had been reported in previous ADHD GWAS studies (Lasky-Su and others 2008), (Neale and others 2010a), (Zhou and others 2008).</p>



SCN10A: *chr3: 38787799-38889198*
GJA1: *rs7753979*
TNNT2: *chr1: 199488134-199580414*

SCN10A: Voltage-gated sodium channels are integral membrane glycoproteins that are responsible for the initial rising phase of action in most excitable cells.

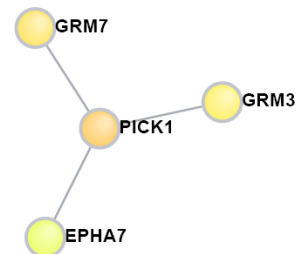
GJA1: This gene is a member of the connexin gene family. The encoded protein is a component of gap junctions. Gap channels allow electrical and biochemical coupling between cells and in excitable tissues, such as neurons and heart.

TNNT2: The protein encoded by this gene is the tropomyosin -binding subunit of the troponin complex, which is located on the thin filament of striated muscles and regulates muscle contraction in response to alterations in intracellular calcium ion concentration. This gene expresses highest in the heart, but also expresses in the brain.

4 SNPs of *SCN10A* showed nominal association with ADHD in the meta-analysis by Neale et al. (2010), with the lowest p value =0.022 for rs7430438.

rs7740467, which is approximately 3kb upstream of *GJA1*, was found nominal significant (p=0.0204) in the meta-analysis of ADHD GWAS by Neale, et al. (2010).

8 SNPs of *TNNT2* showed nominal association with ADHD in the meta-analysis by Neale et al. (2010), with the lowest p value =0.015 for rs10800775



GRM7: *rs13317247*
GRM3: *chr7: 86132321-86244019*
PICK1: *rs8142185*
EPHA7: *chr6: 94194161-94216651*

GRM7 and GRM3: L-glutamate is the major excitatory neurotransmitter in the central nervous system, and it activates both ionotropic and metabotropic glutamate receptors. Glutamatergic neurotransmission is involved in most aspects of normal brain function. ***GRM7 and GRM3***: L-glutamate is the major excitatory neurotransmitter in the central nervous system, and it activates both ionotropic and metabotropic glutamate receptors function and can be perturbed in

More than 20 SNPs of ***GRM7*** showed association with ADHD in the meta-analysis by Neale et al. (2010), with the lowest p value =2.96E-3 for rs1532544. CNVs of ***GRM7*** were found in a Genome-wide copy number variation study of ADHD (Elia and others 2012). rs17031835, rs12491620, rs1450099, and rs3749380 of ***GRM7*** were associated with SCZ (Ganda and others 2009; Ohtsuki and others 2008; Shibata and others 2009).

many neuropathologic conditions. GRM3 belongs to the metabotropic glutamate receptors Group II, while GRM7 belongs to Group III.

PICK1: The protein encoded by this gene has been shown to interact with multiple glutamate receptor subtypes, Probable adapter protein that bind to and organize the subcellular localization of a variety of membrane proteins containing some PDZ recognition sequence. Involved in the clustering of various receptors, possibly by acting at the receptor internalization level. Plays a role in synaptic plasticity by regulating the trafficking and internalization of AMPA receptors.

EPHA7: This gene belongs to the ephrin receptor subfamily of the protein-tyrosine kinase family. EPH and EPH-related receptors have been implicated in mediating developmental events, particularly in the nervous system.

GRM3 and EPHA3 were identified to be candidate genes for ASD (Casey and others 2012). **GRM3** was associated with SCZ in some candidate gene studies (Cherlyn and others 2010). It was also related to [psychosis and relapse in bipolar disorder](#) (Dalvie and others 2010).

PICK1 Located in the linkage and association region of SCZ (Fujii and others 2006; Hong and others 2004; Pulver and others 1994).

rs12661215 of **EPHA7** was found nominal significant ($p=0.0107$) in the meta-analysis of ADHD GWAS by Neale, et al. (2010).

rs2664283 in **CAMK2G** was found nominal significant ($p=7.5E-3$) in the meta-analysis of ADHD GWAS by Neale, et al. (2010)

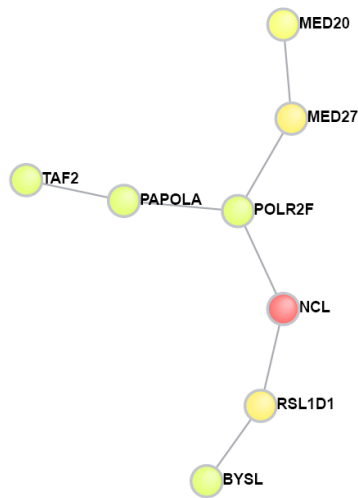
Metabotropic glutamate receptor genes family and their interacting genes were previously found to be enriched with CNVs in ADHD samples (Elia and others 2012).



CAMK2G: rs10824051,
rs11000831
ADAM28: rs7012077

CAMK2G: The product of this gene is one of the four subunits of an enzyme which belongs to the Ca(2+)/ calmodulin- dependent protein kinase subfamily. Calcium signalling is crucial for several aspects of plasticity at glutamatergic synapses.

ADAM28: This gene encodes a member of the ADAM (a disintegrin and metalloprotease domain) family. Members of this family have been implicated in a variety of biological processes involving cell-cell and cell-matrix interactions, including neurogenesis.



MED20: chr6: 41904795 42015370
MED27: rs10512416, rs6597539
TAF2: rs3812463, rs6469849, rs6469852,
rs6989791, rs7012857
PAPOLA: rs7149784, rs7160641
POLR2F: rs8142185
NCL: rs16828074
RSL1D1: chr16: 11823863-11943065
BYSL: chr6: 41904795-42015370

MED20: Component of the Mediator complex, a co-activator involved in the regulated transcription of nearly all RNA polymerase II-dependent genes.

MED27: mediator complex subunit 27. These factors work with co-activators to direct transcriptional initiation by the RNA polymerase II apparatus.

TAF2: stabilizes TFIID binding to core promoter. Transcription factor TFIID is one of the general factors required for accurate and regulated initiation by RNA polymerase II.

PAPOLA: Polymerase that creates the 3'-poly(A) tail of mRNA's.

POLR2F: This gene encodes the sixth largest subunit of RNA polymerase II, the polymerase responsible for synthesizing messenger RNA in eukaryotes, that is also shared by the other two DNA-directed RNA polymerases.

NCL: Nucleolin. play a role in pre-rRNA transcription and ribosome assembly. May play a role in the process of transcriptional elongation.

RSL1D1: ribosomal L1 domain containing

BYSL: Required for processing of 20S pre-rRNA precursor and biogenesis of 40S ribosomal subunits. May be required for regulating cell adhesion during implantation of human embryos

rs3218100, 15kb downstream of **MED20**, was nominal significant ($p=7.06E-3$) in the meta-analysis of ADHD GWAS by Neale, et al. (2010).

rs10901091, which is approximately 90kb upstream of **MED27**, was found nominal significant ($p=1.13E-3$) in the meta-analysis of ADHD GWAS by Neale, et al. (2010). 3 SNPs, approximately 135kb downstream of **PAPOLA**, showed association with ADHD in the meta-analysis by Neale et al. (2010), with the lowest p value = $7.02E-3$ for rs1969795.

POLR2F Located in the linkage and association region of SCZ (Fujii and others 2006; Hong and others 2004; Pulver and others 1994). **RSL1D1** located in 16p13 of the linkage region for ADHD (Fisher and others 2002; Smalley and others 2002). A SNP, rs464017, 65kb downstream of RSL1D1, was nominal significant ($p=0.0387$) in the meta-analysis of ADHD GWAS by Neale, et al. (2010). A SNP, rs3218100, about 4kb downstream of **BYSL**, was nominal significant ($p=7.06E-3$) in the meta-analysis of ADHD GWAS by Neale, et al. (2010).

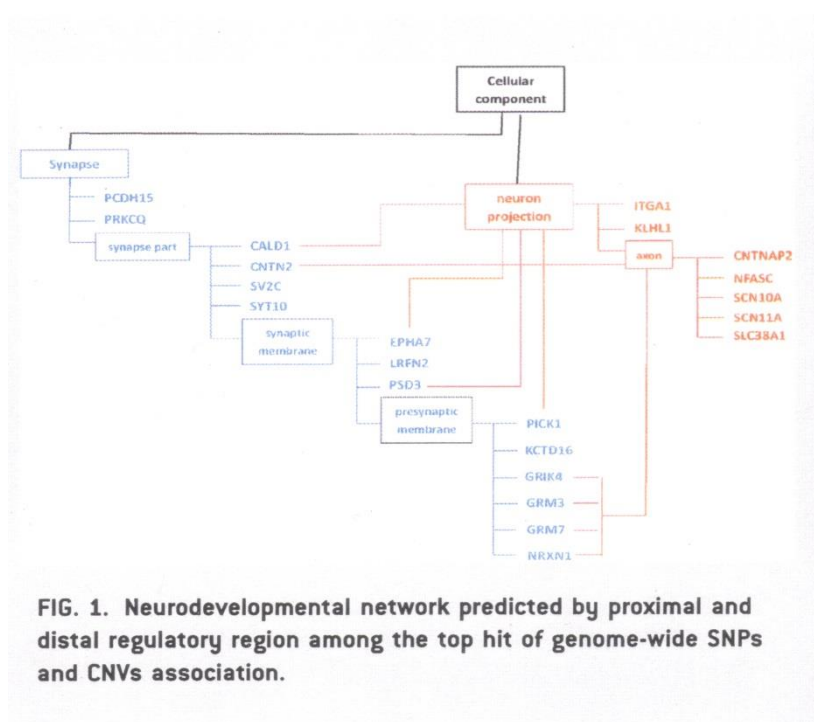
Transcription regulating proteins, such as members of *ZNF* family, *DMRT2*, *FHIT*, *FOXP1*, and *MEIS2*, had been implicated in previous GWAS (Lasky-Su and others 2008; Neale and others 2010b).

TABLE III
Significant enriched gene ontology terms by genes associated with regulatory regions

Term name	Binom* rank	Binom raw P	Binom FDR Q-Val	Binom fold enrichment	Binom observed Region hits	Binom region set coverage %
Presynaptic Membrane	1	2.4669 ⁻¹¹	2.7654 ⁻⁸	9.4056	16	10.53
Synaptic Membrane	2	4.2028 ⁻¹¹	2.3557 ⁻⁸	4.7787	26	17.11
Synapse part	4	1.2576 ⁻⁹	3.5243 ⁻⁷	3.5619	30	19.74
Synapse	5	6.4704 ⁻⁸	1.4507 ⁻⁵	2.8497	32	21.05
Axon	11	4.2006 ⁻⁵	4.2808 ⁻³	2.9590	18	11.84
Neuron projection	13	1.5147 ⁻⁴	1.3062 ⁻²	2.0886	28	18.42

	Hyper Rank	Hyper FDR Q-Val	Hyper fold enrichment	Hyper observed gene hits	Hyper total genes	Hyper gene set coverage %
Presynaptic Membrane	1	5.6208 ⁻³	13.5654	6	50	3.85
Synaptic Membrane	5	1.3568 ⁻²	5.2175	9	195	5.77
Synapse part	2	6.9311 ⁻³	4.2597	13	345	8.33
Synapse	3	5.4656 ⁻³	3.6862	15	460	9.62
Axon	7	1.4342 ⁻²	4.4506	10	254	6.41
Neuron Projection	4	1.2807 ⁻²	3.1788	16	569	10.26

“Binom” represented binomial test over genomic regions. “Hyper” represented hypergeometric test over genes



data, we also used DAPPLE, which is based only on experimentally documented physical interactions among proteins. Considering the complexity of the genetic basis of ADHD, we hypothesized that both common and rare variants contribute to the disorder and act on similar functional classes of genes [Poelmans et al., 2011; Stergiakouli et al., 2012]. The DAPPLE analyses showed that the proteins implicated by our GWAS were significantly more likely to be interconnected with one another than expected by chance, suggesting that risk variants might exist in suites of genes involved in the underlying biological process of protein-protein interaction networks. The DAPPLE results are consistent with the INRICH results implicating three pathways: cell adhesion (NXPH1-NRXN1, CNTN2-CNTNAP2-ZMIZ1), glutamate synaptic development (GRM7-PICK1-GRM3, PICK1-EPHA7), and the transcription pathway (TAF2-PAPOLA-POLR2F-MED27-MED20, POLR2F-NCL-RSL1D1-BYSL).

Using the regulatory annotation of associated signals, GREAT depicted a clearer outline of associated genes, which encoded proteins comprising neuronal cellular components from the Neuron Projection and Synapse branches of the GO tree. Most of the genes from these pathways were consistent with the INRICH and DAPPLE findings; they encode adhesion molecules, glutamate receptors and proteins involved in axon and synapse development (Supplementary Table SIII).

All the above pathways are consistent with the hypothesis that mis-wiring of the brain during neurodevelopment might cause ADHD. Similar conclusions were drawn by Lesch et al. [2008] and Franke et al. [2009] based on the findings from existing GWAS, which suggested that neuronal spine formation

and plasticity might underlie the pathophysiology of ADHD. Consistent with these ideas, a recent integration of ADHD GWAS findings found significant evidence for a neurodevelopmental network of directed neurite outgrowth [Poelmans et al., 2011]. Although our findings are consistent with prior work, they also provide evidence for a more comprehensive network, involving neuron migration, neurite outgrowth, neuronal morphogenesis, and synaptic plasticity, especially glutamatergic synaptic development. The glutamate system is a reasonable candidate for ADHD's pathophysiology as glutamate is the major excitatory neurotransmitter in the central nervous system, and regulates the catecholaminergic activity which has been implicated in ADHD by neurobiological [Scassellati et al., 2012] and treatment [Faraone and Glatt, 2010] studies.

Although our findings are intriguing, we have only captured fragments of the puzzle of ADHD's etiology in this study. We could not paint the full picture. Our work must be considered in the context of its limitations. We had no genome-wide significant findings for any single variant, which might be due to the sample size. However, our bioinformatic and pathway analyses found some interesting genes and neurobiological pathways which implicate complex neurodevelopmental network underlying ADHD. Our finding of a significant polygenic component suggests that there are many common SNP variants with small effect sizes that increase the risk for ADHD. Individually, these SNPs will be difficult to detect with currently available sample sizes.

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REFERENCES

- Barkley RA. 1998. Attention-deficit hyperactivity disorder: A handbook for diagnosis and treatment. New York: Guilford Press. xii 628 p.p.
- Casey JP, Magalhaes T, Conroy JM, Regan R, Shah N, Anney R, Shields DC, Abrahams BS, Almeida J, Bacchelli E, et al. 2012. A novel approach of homozygous haplotype sharing identifies candidate genes in autism spectrum disorder. *Hum Genet* 131(4):565–579.
- Cherlyn SY, Woon PS, Liu JJ, Ong WY, Tsai GC, Sim K. 2010. Genetic association studies of glutamate, GABA and related genes in schizophrenia and bipolar disorder: A decade of advance. *Neurosci Biobehav Rev* 34(6):958–977.

- Dalvie S, Horn N, Nossek C, van der Merwe L, Stein DJ, Ramesar R. 2010. Psychosis and relapse in bipolar disorder are related to GRM3, DAOA, and GRIN2B genotype. *Afr J Psychiatry (Johannesbg)* 13(4):297–301.
- Doherty JL, O'Donovan MC, Owen MJ. 2012. Recent genomic advances in schizophrenia. *Clin Genet* 81(2):103–109.
- Elia J, Gai X, Xie HM, Perin JC, Geiger E, Glessner JT, D'Arcy M, de Berardinis R, Frackelton E, Kim C, et al. 2010. Rare structural variants found in attention-deficit hyperactivity disorder are preferentially associated with neurodevelopmental genes. *Mol Psychiatry* 15(6):637–646.
- Elia J, Glessner JT, Wang K, Takahashi N, Shtir CJ, Hadley D, Sleiman PM, Zhang H, Kim CE, Robison R, et al. 2012. Genome-wide copy number variation study associates metabotropic glutamate receptor gene networks with attention deficit hyperactivity disorder. *Nat Genet* 44(1): 78–84.
- Faraone SV, Glatt SJ. 2010. A comparison of the efficacy of medications for adult attention-deficit/hyperactivity disorder using meta-analysis of effect sizes. *J Clin Psychiatry* 71(6):754–763.
- Faraone SV, Sergeant J, Gillberg C, Biederman J. 2003. The worldwide prevalence of ADHD: Is it an American condition? *World Psychiatry* 2 (2):104–113.
- Fisher SE, Francks C, McCracken JT, McGough JJ, Marlow AJ, MacPhie IL, Newbury DF, Crawford LR, Palmer CG, Woodward JA, et al. 2002. A genome-wide scan for loci involved in attention-deficit/hyperactivity disorder. *Am J Hum Genet* 70(5):1183–1196.
- Franke B, Neale BM, Faraone SV. 2009. Genome-wide association studies in ADHD. *Hum Genet* 126(1):13–50.
- Fujii K, Maeda K, Hikida T, Mustafa AK, Balkissoon R, Xia J, Yamada T, Ozeki Y, Kawahara R, Okawa M, et al. 2006. Serine racemase binds to PICK1: Potential relevance to schizophrenia. *Mol Psychiatry* 11(2): 150–157.
- Ganda C, Schwab SG, Amir N, Heriani H, Irmansyah I, Kusumawardhani A, Nasrun M, Widyawati I, Maier W, Wildenauer DB. 2009. A family based association study of DNA sequence variants in GRM7 with schizophrenia in an Indonesian population. *Int J Neuropsychopharmacol* 12(9):1283–1289.
- Hong CJ, Liao DL, Shih HL, Tsai SJ. 2004. Association study of PICK1 rs3952 polymorphism and schizophrenia. *Neuroreport* 15(12):1965–1967.
- Lasky-Su J, Neale BM, Franke B, Anney RJ, Zhou K, Maller JB, Vasquez AA, Chen W, Asherson P, Buitelaar J, et al. 2008. Genome-wide association scan of quantitative traits for attention deficit hyperactivity disorder identifies novel associations and confirms candidate gene associations. *Am J Med Genet Part B* 147B(8):1345–1354.
- Lee SH, Wray NR, Goddard ME, Visscher PM. 2011. Estimating missing heritability for disease from genome-wide association studies. *Am J Hum Genet* 88(3):294–305.
- Lee PH, O'Dushlaine C, Thomas B, Purcell SM. 2012a. INRICH: Interval based enrichment analysis for genome wide association studies. *Bioinformatics* 28(13):1797–1799.
- Lee SH, Yang J, Goddard ME, Visscher PM, Wray NR. 2012b. Estimation of pleiotropy between complex diseases using single-nucleotide polymorphism-derived genomic relationships and restricted maximum likelihood. *Bioinformatics* 28(19):2540–2542.
- Lesch KP, Timmesfeld N, Renner TJ, Halperin R, Roser C, Nguyen TT, Craig DW, Romanos J, Heine M, Meyer J, et al. 2008. Molecular genetics of adult ADHD:

- Converging evidence from genome-wide association and extended pedigree linkage studies. *J Neural Transmembr* 115 (11):1573–1585.
- McLean CY, Bristor D, Hiller M, Clarke SL, Schaar BT, Lowe CB, Wenger AM, Bejerano G. 2010. GREAT improves functional interpretation of cis-regulatory regions. *Nat Biotechnol* 28(5):495–501.
- Neale BM, Lasky-Su J, Anney R, Franke B, Zhou K, Maller JB, Vasquez AA, Asherson P, Chen W, Banaschewski T, et al. 2008. Genome-wide association scan of attention deficit hyperactivity disorder. *Am J Med Genet Part B* 147B (8):1337–1344.
- Neale BM, Medland S, Ripke S, Anney RJ, Asherson P, Buitelaar J, Franke B, Gill M, Kent L, Holmans P, et al. 2010a. Case-control genome-wide association study of attention-deficit/hyperactivity disorder. *J Am Acad Child Adolesc Psychiatry* 49(9):906–920.
- Neale BM, Medland SE, Ripke S, Asherson P, Franke B, Lesch KP, Faraone SV, Nguyen TT, Schafer H, Holmans P, et al. 2010b. Meta-analysis of genome-wide association studies of attention-deficit/hyperactivity disorder. *J Am Acad Child Adolesc Psychiatry* 49(9):884–897.
- Ohtsuki T, Koga M, Ishiguro H, Horiuchi Y, Arai M, Niizato K, Itokawa M, Inada T, Iwata N, Iritani S, et al. 2008. A polymorphism of the metabotropic glutamate receptor mGluR7 (GRM7) gene is associated with schizophrenia. *Schizophr Res* 101(1–3):9–16.
- Poelmans G, Pauls DL, Buitelaar JK, Franke B. 2011. Integrated genome-wide association study findings: identification of a neurodevelopmental network for attention deficit hyperactivity disorder. *Am J Psychiatry* 168 (4):365–377.
- Pulver AE, Karayiorgou M, Wolyniec PS, Lasseter VK, Kasch L, Nestadt G, Antonarakis S, Housman D, Kazazian HH, Meyers D, et al. 1994. Sequential strategy to identify a susceptibility gene for schizophrenia: report of potential linkage on chromosome 22q12-q13.1: Part 1. *Am J Med Genet* 54(1):36–43.
- Purcell S, Neale B, Todd-Brown K, Thomas L, Ferreira MA, Bender D, Maller J, Sklar P, de Bakker PI, Daly MJ, et al. 2007. PLINK: A tool set for whole-genome association and population-based linkage analyses. *Am J Hum Genet* 81(3):559–575.
- Rossin EJ, Lage K, Raychaudhuri S, Xavier RJ, Tatar D, Benita Y, International Inflammatory Bowel Disease Genetics C, Cotsapas C, Daly MJ. 2011. Proteins encoded in genomic regions associated with immune mediated disease physically interact and suggest underlying biology. *PLoS Genet* 7(1):e1001273.
- Salyakina D, Cukier HN, Lee JM, Sacharow S, Nations LD, Ma D, Jaworski JM, Konidari I, Whitehead PL, Wright HH, et al. 2011. Copy number variants in extended autism spectrum disorder families reveal candidates potentially involved in autism risk. *PLoS ONE* 6(10):e26049.
- Scassellati C, Bonvicini C, Faraone SV, Gennarelli M. 2012. Biomarkers and attention-deficit/hyperactivity disorder: A systematic review and meta-analyses. *J Am Acad Child Adolesc Psychiatry* 51(10):1003–1019; e20.
- Shibata H, Tani A, Chikuhara T, Kikuta R, Sakai M, Ninomiya H, Tashiro N, Iwata N, Ozaki N, Fukumaki Y. 2009. Association study of polymorphisms in the group III metabotropic glutamate receptor genes, GRM4 and GRM7, with schizophrenia. *Psychiatry Res* 167(1–2): 88–96.
- Smalley SL, Kustanovich V, Minassian SL, Stone JL, Ogdie MN, McGough JJ, McCracken JT, MacPhie IL, Francks C, Fisher SE, et al. 2002. Genetic linkage of attention-deficit/hyperactivity disorder

- on chromosome 16p13, in a region implicated in autism. *Am J Hum Genet* 71(4):959–963.
- Stergiakouli E, Hamshere M, Holmans P, Langley K, Zaharieva I, Hawi Z, Kent L, Gill M, Williams N, Owen MJ, et al. 2012. Investigating the contribution of common genetic variants to the risk and pathogenesis of ADHD. *Am J Psychiatry* 169(2):186–194.
- Tsiotra PC, Karageorgos D, Theodorakis K, Michaelidis TM, Modi WS, Furley AJ, Jessell TM, Papamatheakis J. 1993. Isolation of the cDNA and chromosomal localization of the gene (TAX1) encoding the human axonal glycoprotein TAG-1. *Genomics* 18(3):562–567.
- Williams NM, Zaharieva I, Martin A, Langley K, Mantripragada K, Fossdal R, Stefansson H, Stefansson K, Magnusson P, Gudmundsson, et al. 2010. Rare chromosomal deletions and duplications in attention-deficit hyperactivity disorder: A genome-wide analysis. *Lancet* 376(9750):1401–1408.
- Williams NM, Franke B, Mick E, Anney RJ, Freitag CM, Gill M, Thapar A, O'Donovan MC, Owen MJ, Holmans P, et al. 2012. Genome-wide analysis of copy number variants in attention deficit hyperactivity disorder: the role of rare variants and duplications at 15q13.3. *Am J Psychiatry* 169(2):195–204.
- Yang L, Wang YF, Qian QJ, Biederman J, Faraone SV. 2004. DSM-IV subtypes of ADHD in a Chinese outpatient sample. *J Am Acad Child Adolesc Psychiatry* 43(3):248–250.
- Yang J, Benyamin B, McEvoy BP, Gordon S, Henders AK, Nyholt DR, Madden PA, Heath AC, Martin NG, Montgomery GW, et al. 2010. Common SNPs explain a large proportion of the heritability for human height. *Nat Genet* 42:565–569.
- Yang J, Lee SH, Goddard ME, Visscher PM. 2011. GCTA: A tool for genome-wide complex trait analysis. *Am J Hum Genet* 88(1): 76–82.
- Zhou K, Dempfle A, Arcos-Burgos M, Bakker SC, Banaschewski T, Biederman J, Buitelaar J, Castellanos FX, Doyle A, Ebstein RP, et al. 2008. Meta-analysis of genome-wide linkage scans of attention deficit hyperactivity disorder. *Am J Med Genet Part B* 147B (8): 1392–1398.

SUPPORTING INFORMATION

Additional supporting information may be found in the online *version of this article* at the publisher's web-site.

FIG. S1. Pair-wise multi-dimensional scaling (MDS) for 10 dimensions.

FIG. S2. Quantile–quantile(QQ)plot for GWAS of ADHD in a Han Chinese case-control sample.

FIG. S3. Manhattan plot for GWAS of ADHD in a Han Chinese case-control sample (corrected by principal components analysis for genomic background).

FIGURE 1. S4. Direct and indirect networks and prioritized genes from associated SNP and CNV intervals.

TABLE SI. Top Hit SNPs ($P < 10^{-5}$) From Genome-Wide Association Study of ADHD in Han Chinese Case-Control Sample

TABLE SII. Association Analysis of Rare CNV Segments in Cases and Controls

TABLE SIII. Functions of Regulatory Regions Associated Genes That Significantly Enrich GO Terms